

# Identifying the *Cactoblastis cactorum* Pheromone

<sup>†</sup>R. R. Heath, <sup>†</sup>N. D. Epsky, <sup>††</sup>P. E. A. Teal, <sup>††</sup>B. D. Deuben,  
and <sup>†††</sup>J. Cibrián-Tovar

<sup>†</sup>USDA-ARS-SHRS, Miami, FL

<sup>††</sup>USDA-ARS-CMAVE, Gainesville, FL

<sup>†††</sup>Colegio de Postgraduados, Texcoco, Mexico



# Collaborators

**USDA-ARS Subtropical Horticulture Research Station (SHRS, Miami, FL)**

**Paul Kendra**

**USDA-ARS (Tifton, GA; Tallahassee, FL)**

**Jim Carpenter; Stephen Hight**

**USDA-APHIS-PPQ (Raleigh, NC; Riverdale, MD)**

**Ken Bloem, Stephanie Bloem; Joel Floyd**

**International Atomic Energy Agency (IAEA, Austria)**

**Walther Enkerlin, Jorge Hendrichs**

**Colegio de Postgraduados (Texcoco, México)**

**Dirección General de Sanidad Vegetal (SAGARPA, México)**

**Jorge Hernández-Baeza, Arturo Bello**



# Methodology

- gland cuts
- Pheromone Biosynthesis Activating Neuropeptide (PBAN) + gland cuts
- volatile collections



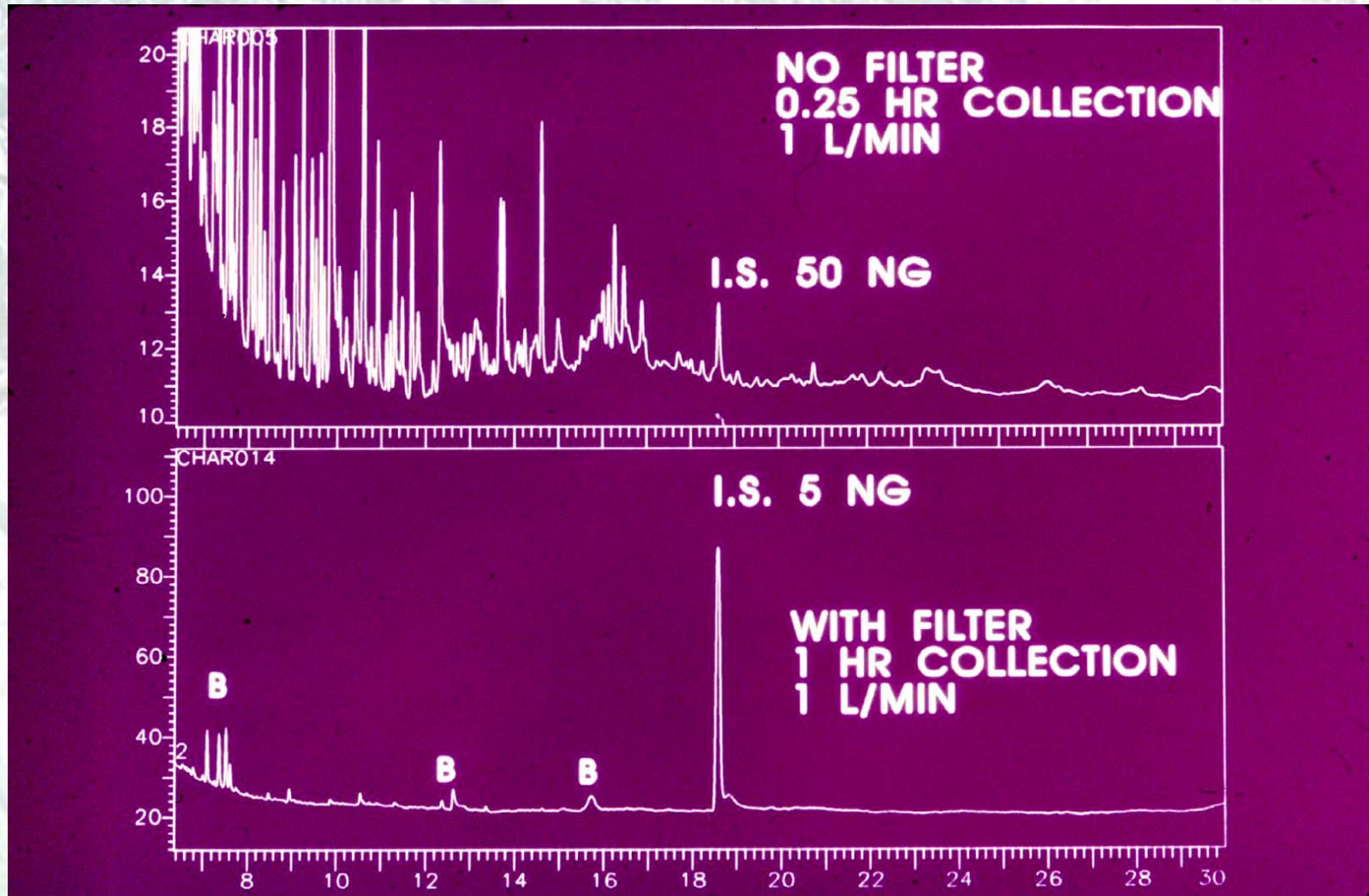
## Preferred system

- a) chemicals identified from gland cuts (with or without PBAN) do not reflect the release rate or ratio of the pheromone system
- b) gland cuts often contain pheromone precursors that may be either not active or inhibitory

Thus, volatile collection is the preferred method

# Systems to collect insect semiochemicals


## Air must be purified and humidified





# Volatile collection apparatus



The background image shows a field of low-growing plants with thick, green, oval-shaped leaves, possibly a coastal or saline species. They are planted in neat rows on a light-colored, sandy or silty soil. The plants are in various stages of growth, with some showing small white flowers. The overall scene is bright and sunny, with some shadows cast by the plants.

**Collections were made  
at 1-2 hour intervals  
From 5:00 a.m. to 8:00 a.m.**



A photograph of a field with rows of green, leafy plants growing in sandy soil. The plants are arranged in neat rows, and the soil is light-colored and appears to be a mix of sand and small pebbles. The background shows more rows of plants extending into the distance under a bright sky.

**Adsorbent traps were  
extracted with  
150  $\mu$ l of methylene chloride**



The background of the slide is a photograph of a field of green, leafy plants, possibly a type of succulent or coastal plant, growing in rows on sandy soil. The plants are small and have thick, rounded leaves. The soil is light-colored and appears to be a mix of sand and small pebbles. The plants are arranged in neat rows, and the overall scene is bright and sunny.

**Prior to analysis,  
the extracted material  
was concentrated**

**Concentration may result in loss of active  
components**

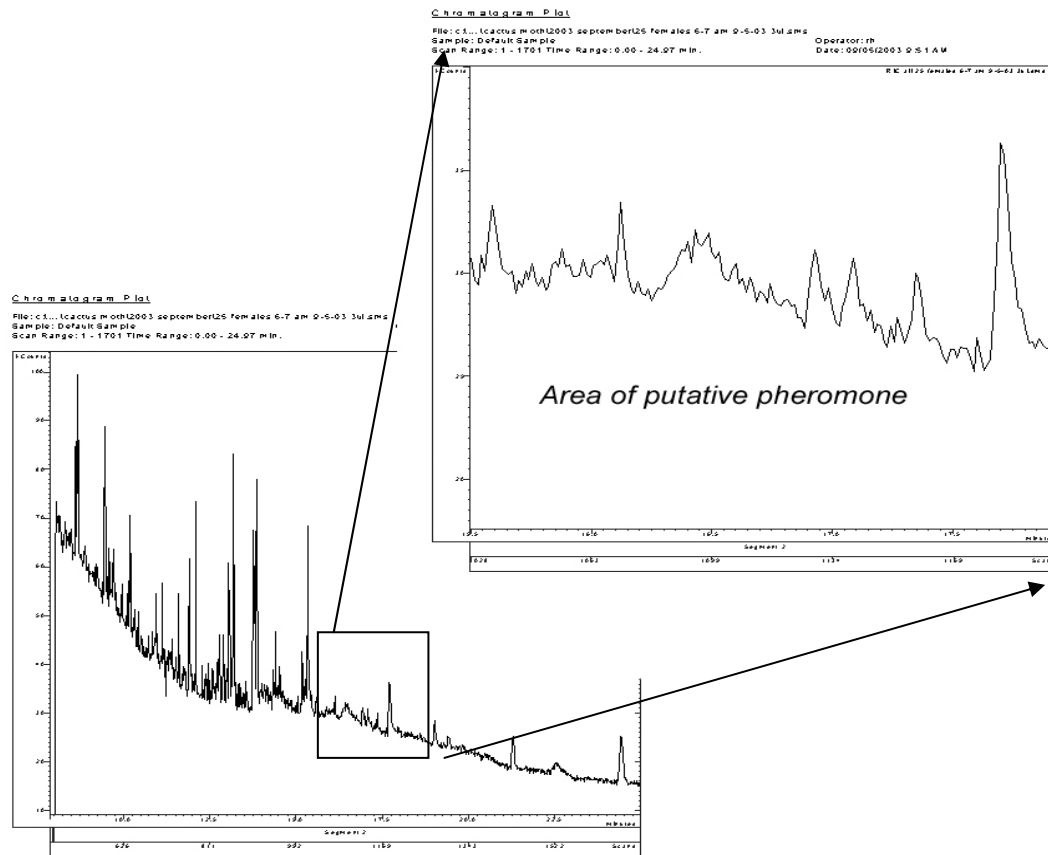
# Identification process

## Using Electron Impact (EI) and Chemical Ionization (CI) with isobutane as the CI reagent





# Current status



Best sample to date: putative compounds  
observed 16 out of 125 times

# Flight tunnel bioassay

Juan Cibrián-Tovar  
Colegio de Postgraduados  
Texcoco, Mexico

Sponsored by the UN, IAEA, Pest Control  
Division, Dr. Jorge Hendrichs and Dr. Walther  
Enkerlin



# Bioassay procedure

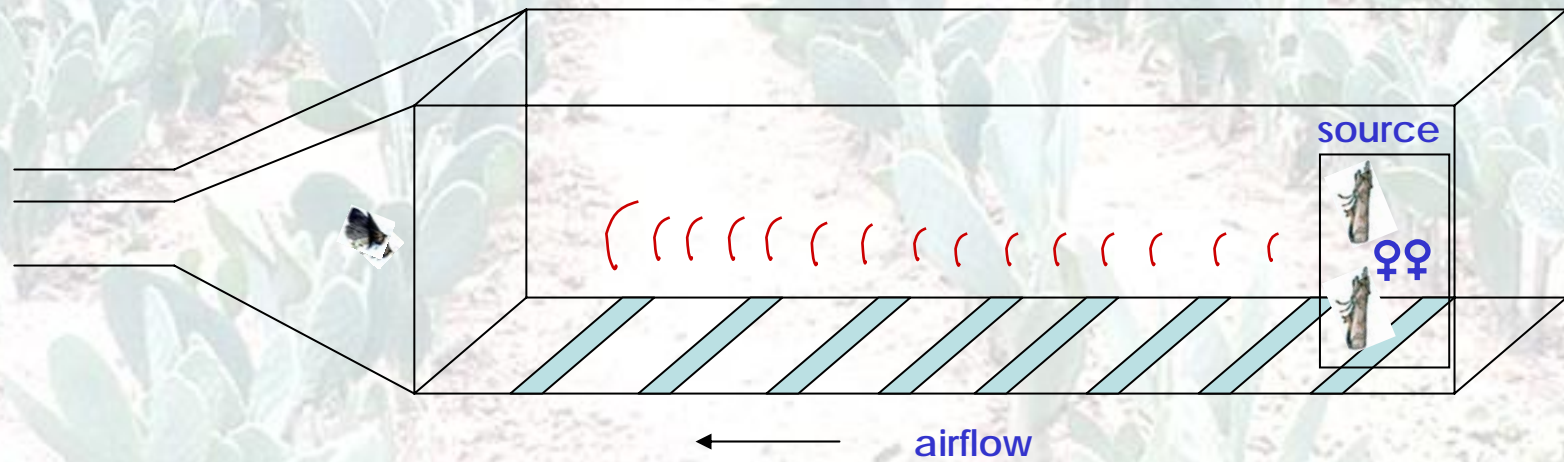
The observations were made in a wind tunnel at 12:12 h photoperiod,  $75 \pm 5$  °F and 80 % RH.

20 males (1-3 days old) were released one by one into the wind tunnel for each treatment.

The treatments were: three live virgin females, the gland extract (3 female equivalents) or the septa with different blends (A, B, C, D).

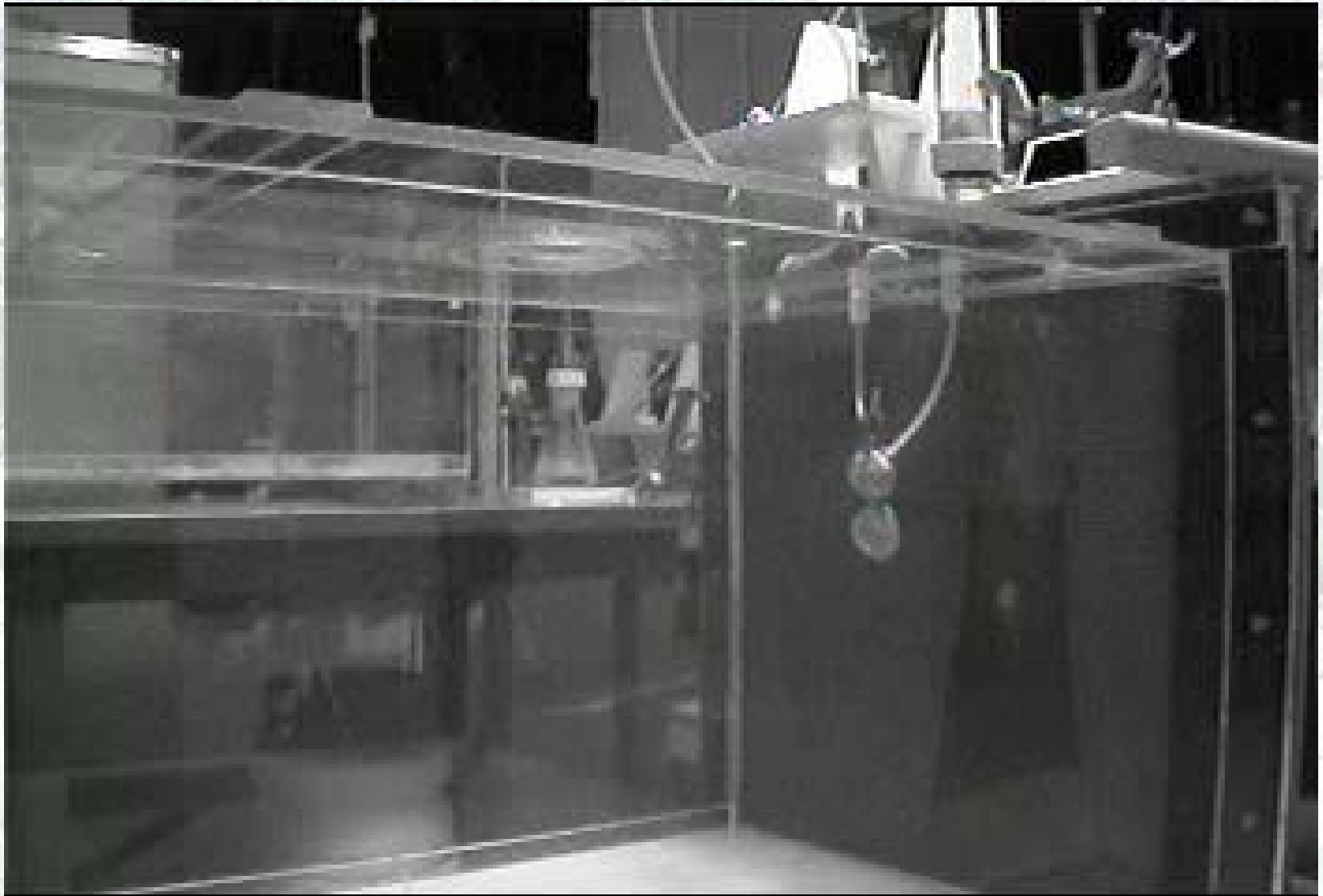
The following behaviors were registered: Oriented Flight (OF) and Landing (LA).

# Flight tunnel animation

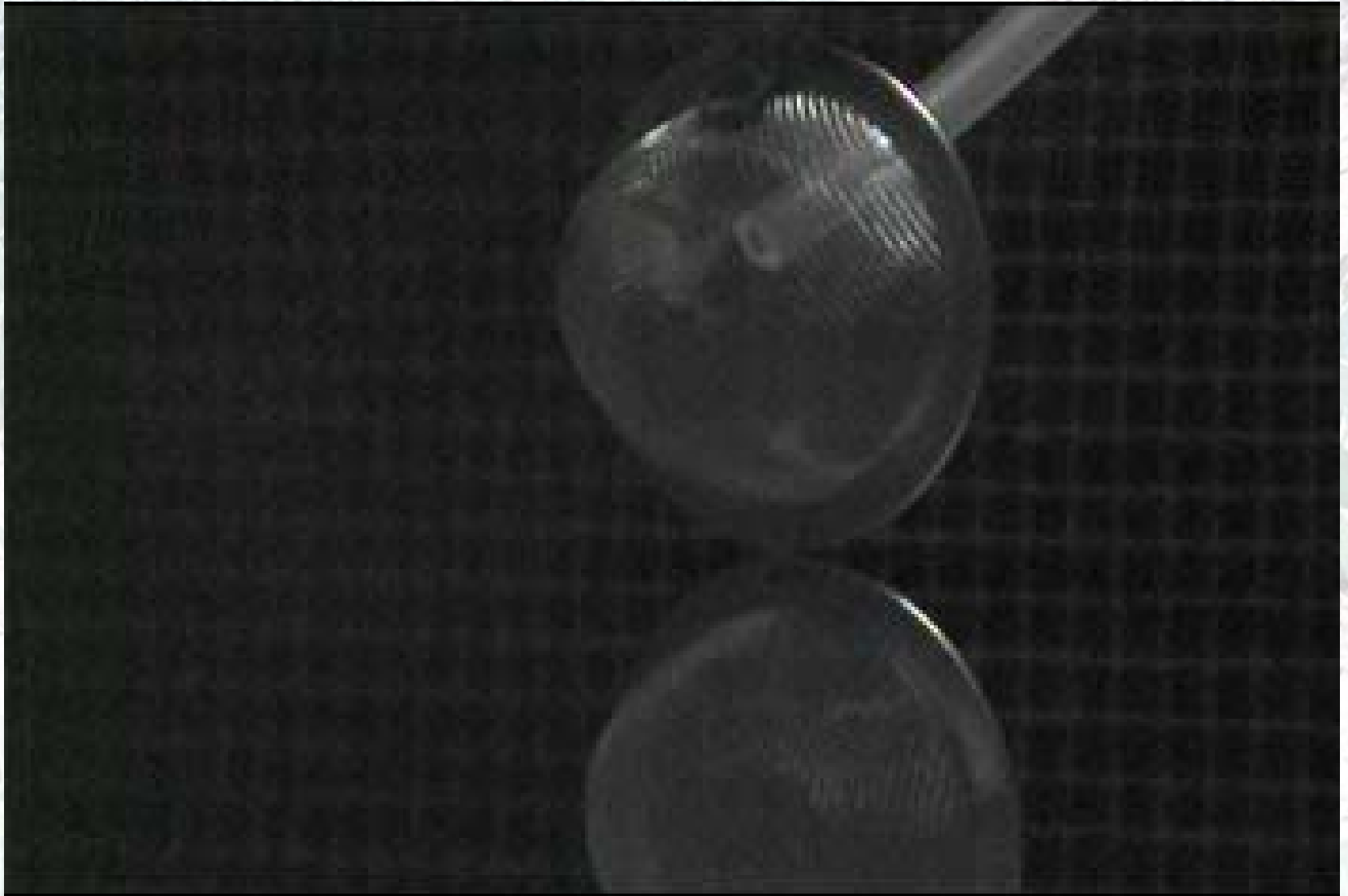




# Flight tunnel – oriented flight

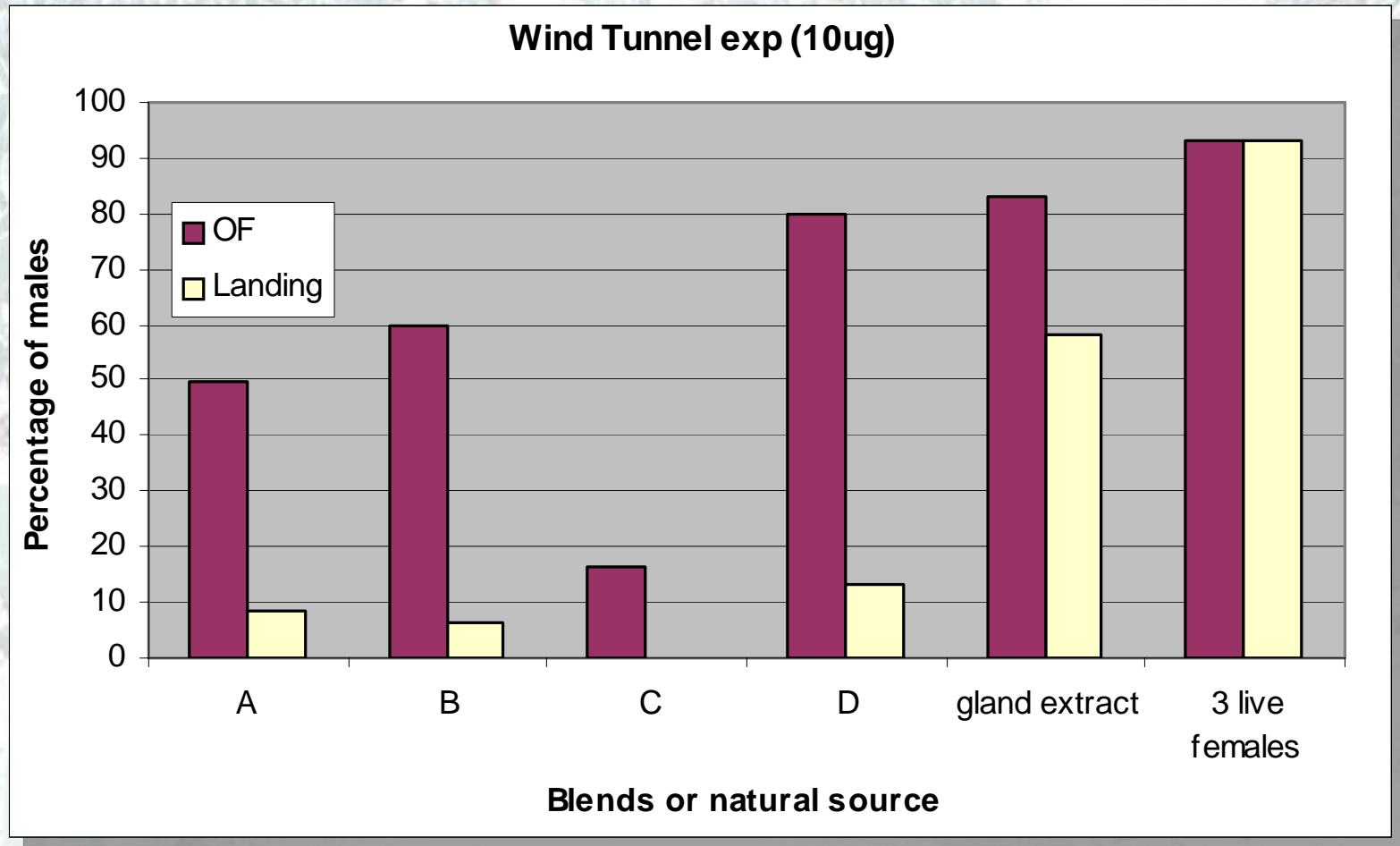


# Flight tunnel - landing





# Bioassay results



# Field Tests

Tests were conducted in collaboration with J. Carpenter, S. Hight, S. Bloem to test capture of cactus moth males in the field.

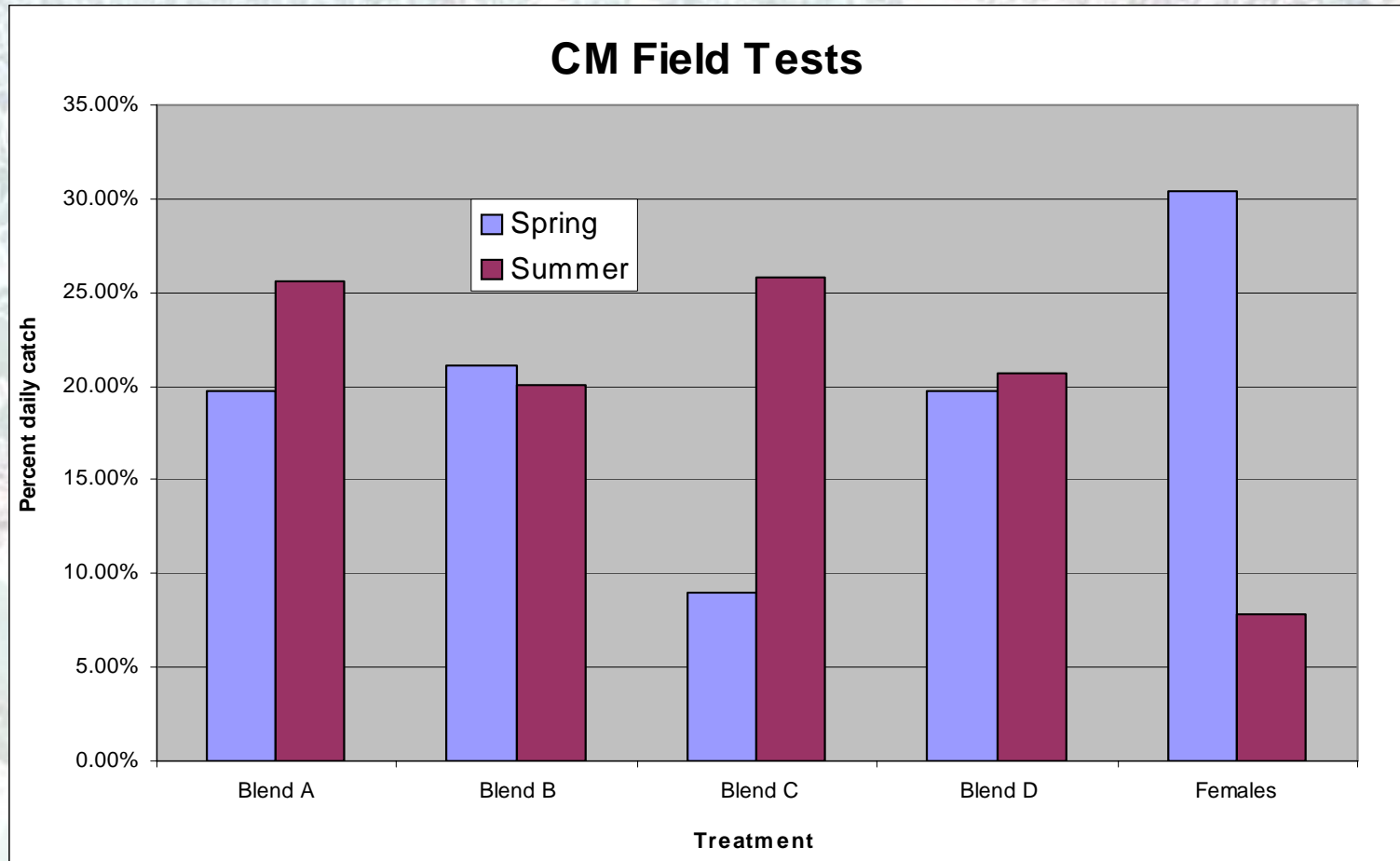
The treatments were C-traps baited with two live virgin females or septa (blend A, B, C or D) prepared by Suterra LLC (Bend, OR).

Tests were run in spring (5 reps, 4.5 wk)  
and summer (5 reps, 5 wk)



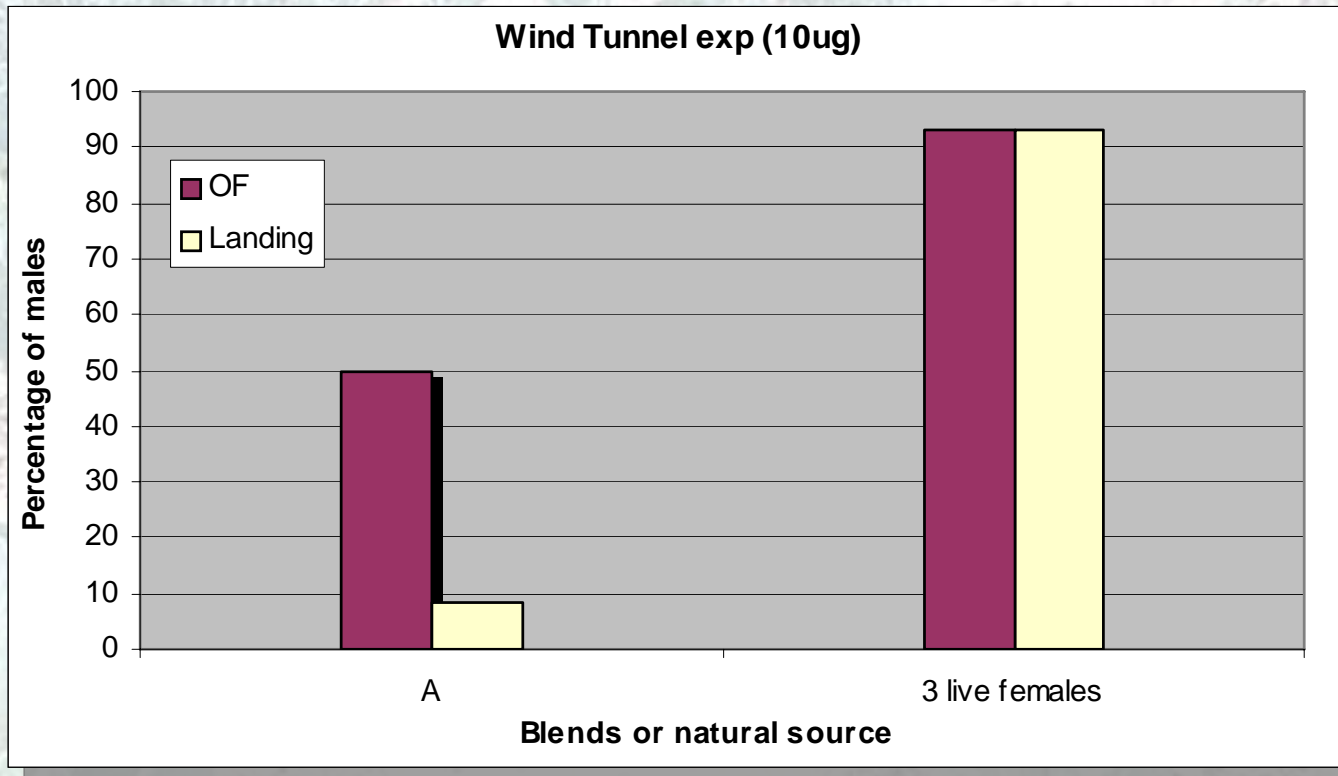


# Field results



**Note: Females are susceptible to environmental conditions...**

# Conclusion



a) Results speak for themselves. We do not have a complete pheromone system developed.

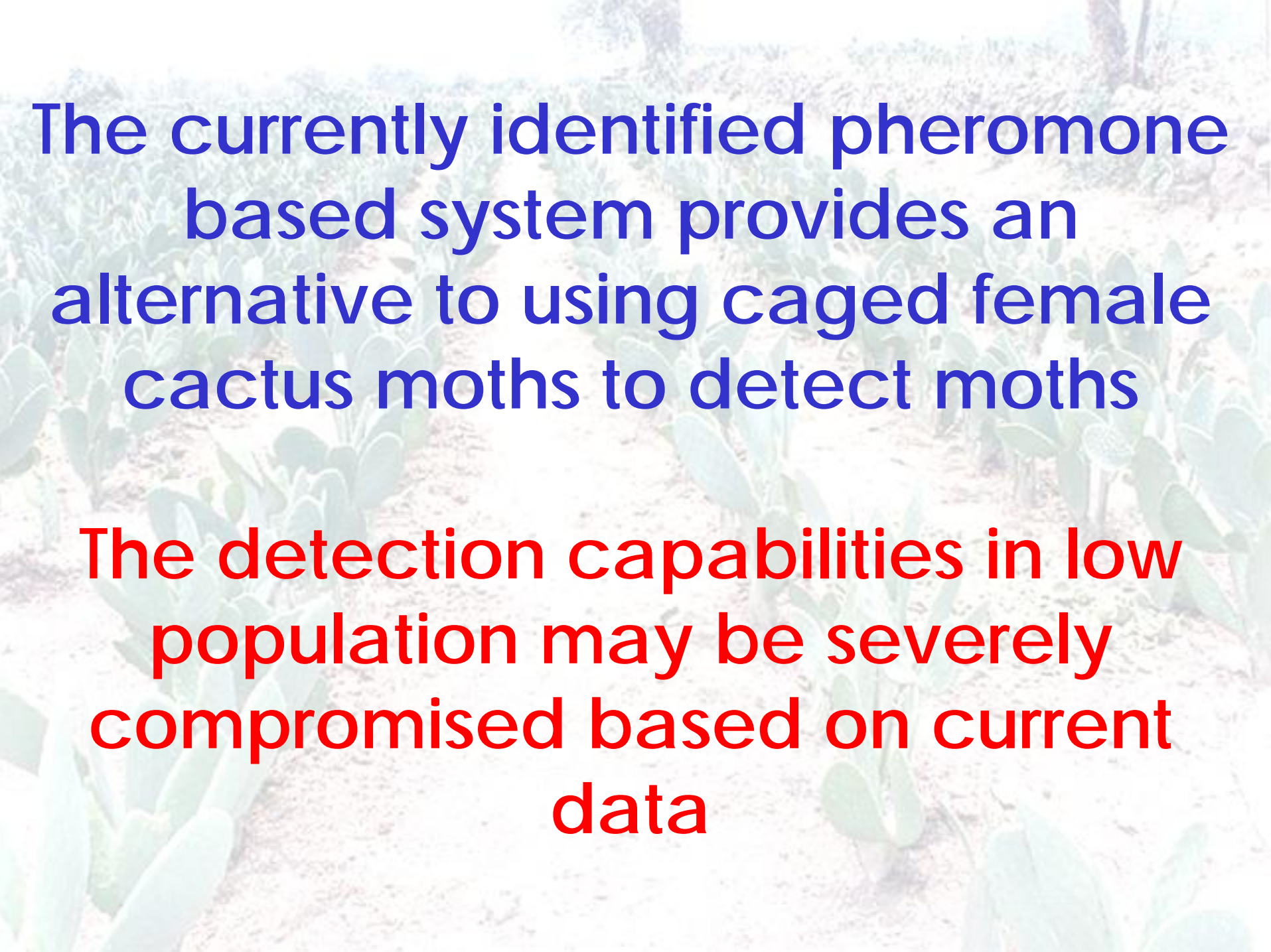
b) Current experimental system used is blend A





**A perspective of  
the current status of the cactus moth  
pheromone system**

**A reality check**



The currently identified pheromone based system provides an alternative to using caged female cactus moths to detect moths

The detection capabilities in low population may be severely compromised based on current data





**Future 2005**

**Identify missing components  
Determine longevity of lures**



**Future 2007**

**Identify missing components  
Determine longevity of lures**



The background of the slide is a photograph of a dirt path or road. On either side of the path are rows of green, leafy plants, possibly a type of shrub or small tree, growing in a field. The path leads into the distance, and the overall scene is brightly lit, suggesting a sunny day.

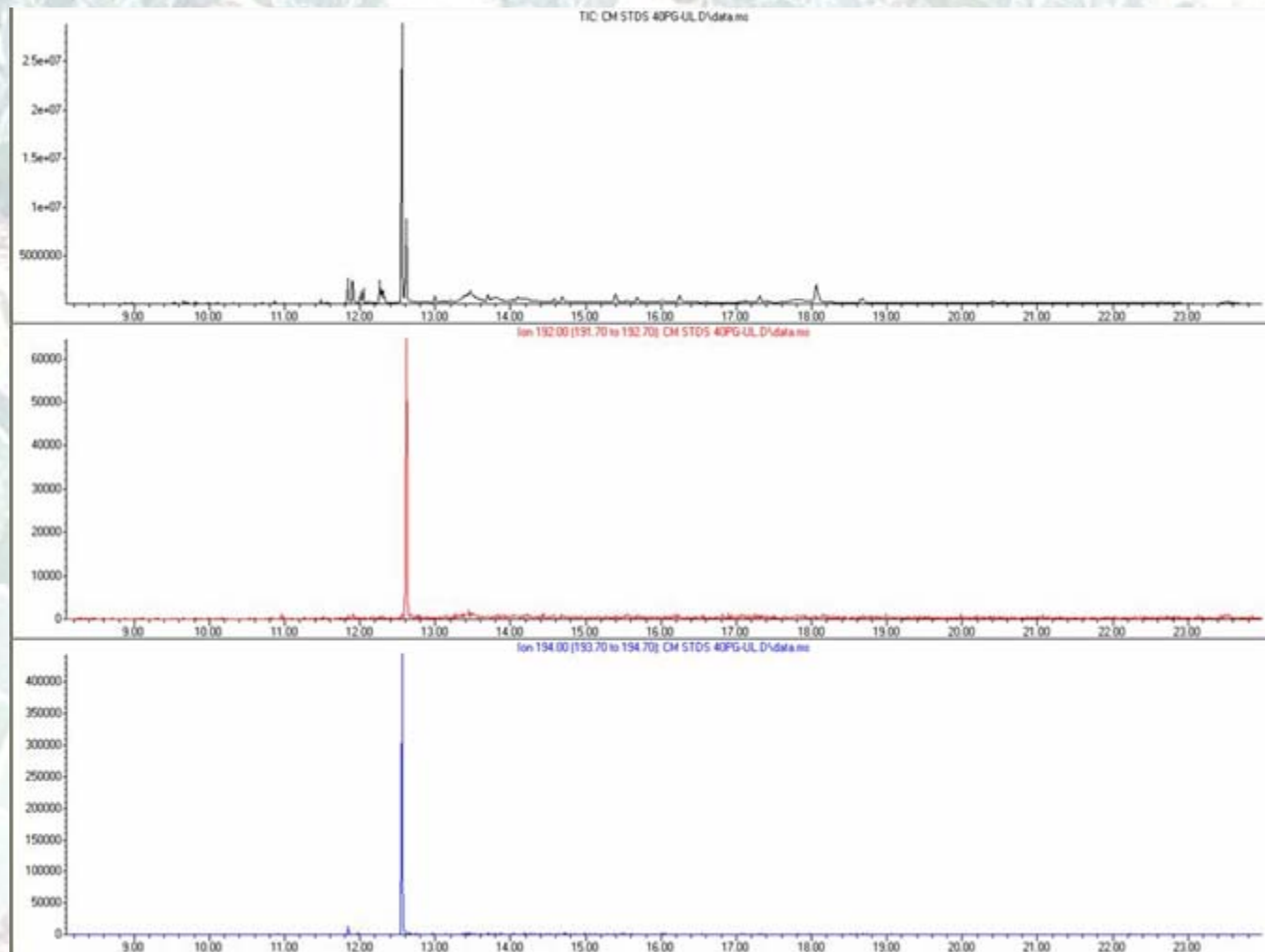
# Not all was lost

## Some significant accomplishments were made

### Good news:

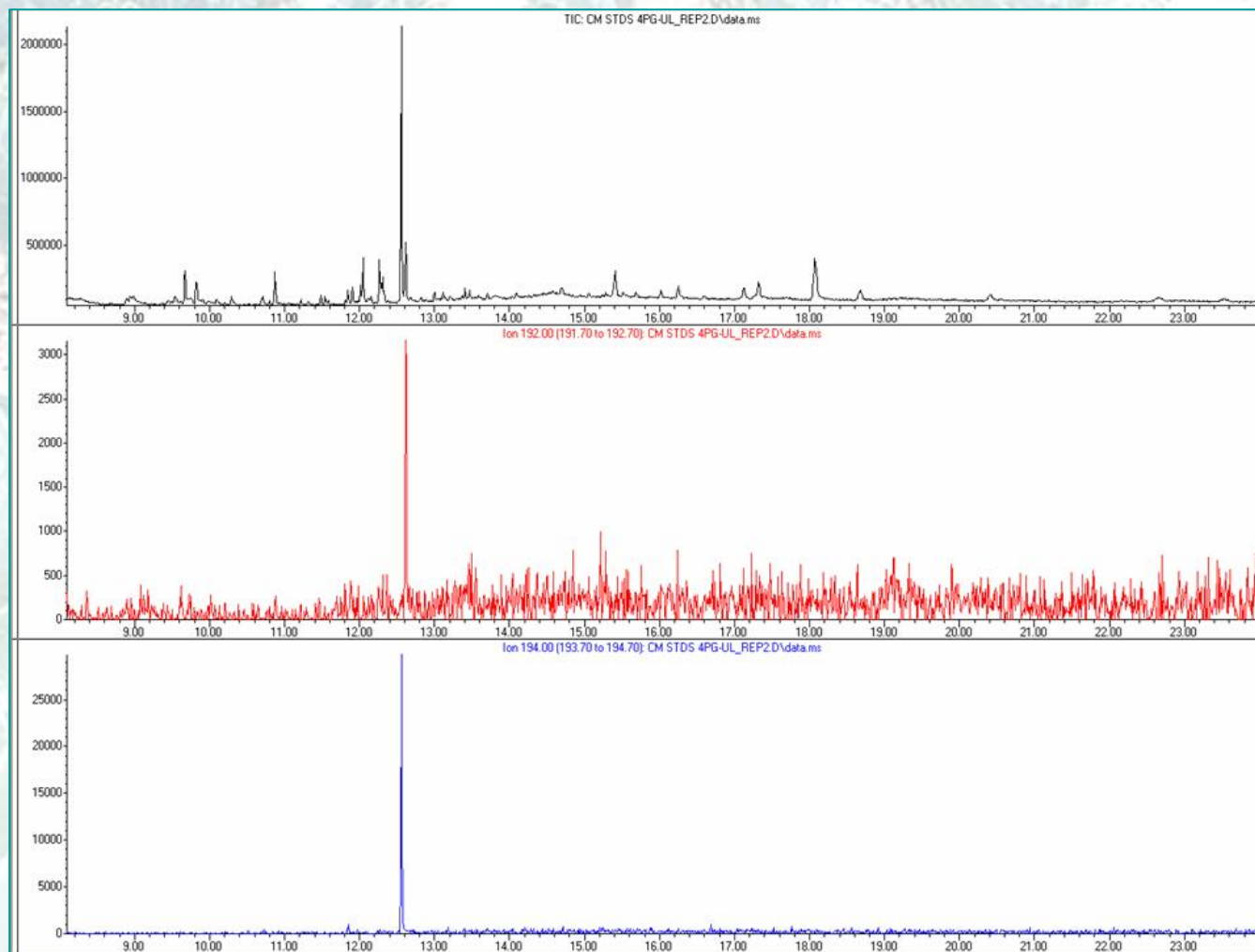
- new GC/MS increased sensitivity of analysis (decrease in amount of material needed)
- previously the amount injected was 2  $\mu\text{L}$ , which required concentrate of sample. Current system permits stackable 25  $\mu\text{L}$  injections so no sample concentration needed

40 pg/uL, 50 uL injected





4 pg/uL, 50 uL injected

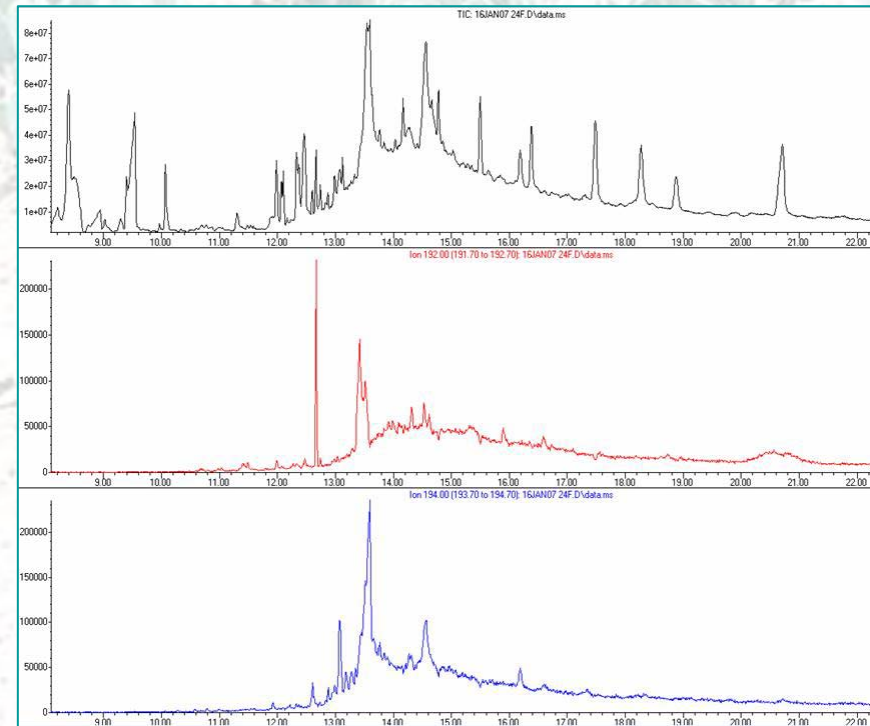
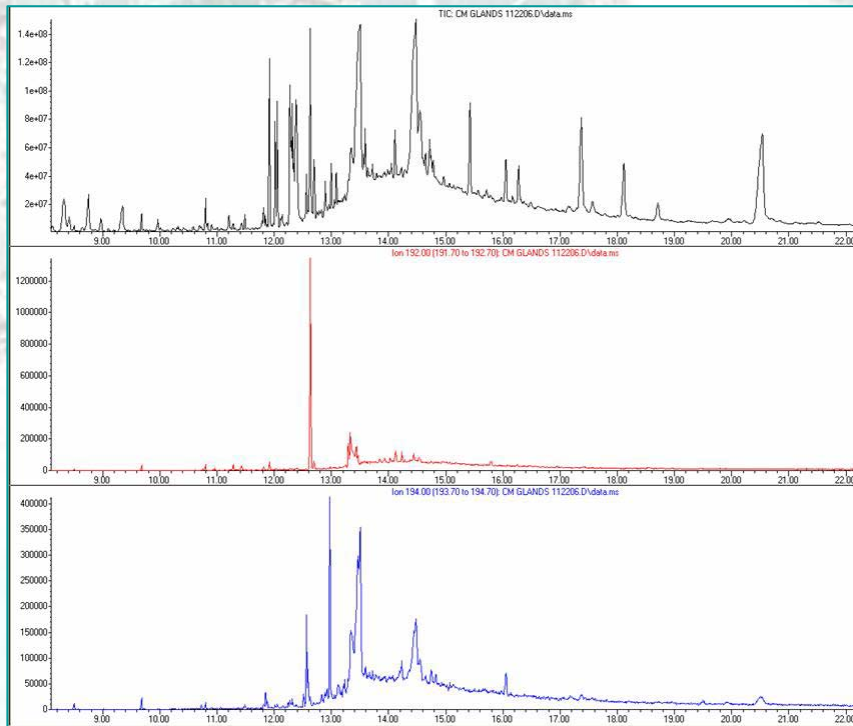


A photograph of a field with rows of green, leafy plants growing in sandy soil. A dirt path runs through the middle of the field, leading towards the background. The text is overlaid in the center of the image.

Oftentimes, with good news,  
you get bad news



Although we obtained a significant increase in detection sensitivity with the new GC/MS, we found that gland extracts obtained in January contained considerably less pheromone than those obtained in October



Currently, we are analyzing gland extracts to see if amounts of materials in moths under spring conditions return to levels observed under fall conditions.

If the expected quantities of pheromone are observed in the glands, volatile chemical collections will be made.

---

If the above fails, a more classical approach may be required. Specifically, this would be to bioassay isolated fractions obtained from gland extracts with the 3 identified compounds.





# Pheromone-Based Attractant for Males of *Cactoblastis cactorum* (Lepidoptera: Pyralidae)

R. R. HEATH, P. E. A. TEAL, N. D. EPSKY, B. D. DUEBEN, S. D. HIGHT, S. BLOEM, J. E. CARPENTER,  
T. J. WEISSLING, P. E. KENDRA, J. CIBRIAN-TOVAR, AND K. A. BLOEM

Environ. Entomol. 35(6): 1469 -1476 (2006)

**ABSTRACT** The cactus moth, *Cactoblastis cactorum* (Berg), is an invasive pest of *Opuntia* spp. Since its arrival in the Florida Keys in 1989, it has moved rapidly up the east and west coasts of Florida, threatening to invade the southwestern United States and Mexico. Female moths produce a sex pheromone that attracts male moths. In this study, we report on mating behavior observed in the laboratory and the identification of putative pheromonal chemical components based on mass spectral analysis of volatiles collected from virgin female moths and from solvent extraction of excised glands. Three candidate components, formulated on rubber septa in different release rates and ratios, were tested in laboratory olfactometer and flight tunnel experiments, and in field tests in areas with known feral populations of cactus moths. Lures formulated with the three-component blend of 54% (*Z,E*)-9,12 tetradecadien-1-ol acetate, 42% (*Z,E*)-9,12 tetradecadien-1-ol, and 4% (*Z*)-9-tetradecen-1-ol acetate were the most effective, although changes in the ratio of these components had little effect on lure efficacy. For field deployment, traps baited with synthetic lures with a 1 mg load of the three component blend captured equal or higher numbers of males than traps baited with two virgin females. Trapping systems using this pheromone-based attractant will be useful for population delineation in areas currently infested.

Paper available for download at [www.entso.org](http://www.entso.org)  
Reprints available here from speaker



# Acknowledgements:

**USDA-ARS (Miami, FL)**

**Paul Kendra, Aimé Vázquez, Elena Schnell, Daniel Mateo,  
Wayne Montgomery, Micah Gill, Alison Walker**

**USDA-ARS (Tifton, GA)**

**Jim Carpenter, Susan Drawdy**

**USDA-ARS (Tallahassee, FL)**

**Stephen Hight, Nathan Herrick**

**USDA-APHIS-PPQ (Raleigh, NC; Riverdale, MD)**

**Ken Bloem, Stephanie Bloem; Joel Floyd**

**International Atomic Energy Agency (IAEA)**

**Walther Enkerlin, Jorge Hendrichs**

**Dirección General de Sanidad Vegetal (SAGARPA, México)**

**Jorge Hernández-Baeza, Arturo Bello**

**Colegio de Postgraduados (Texcoco, México)**

**Suterra LLC (Bend, OR)**

**Joan Fisher**





A photograph of a large field of young prickly pear cacti (Opuntia) planted in neat, parallel rows. The cacti are small and green, with several flat, oval-shaped pads. They are growing in a dry, sandy soil. In the background, there is a low stone wall and some trees, with mountains visible in the distance under a clear sky. The text "THANK YOU" is overlaid in the center of the image.

**THANK YOU**